promoters and the *gpt* promoter to name a few. Other suitable promoters will be known to the skilled artisan.

[0031] The expression constructs will further contain sites for transcription initiation, termination and, in the transcribed region, a ribosome binding site for translation. The coding portion of the mature transcripts expressed by the constructs can include a translation initiating codon at the beginning and a termination codon appropriately positioned at the end of the polypeptide to be translated.

As indicated, the expression vectors will preferably include at least one selectable marker. Preferably the selection marker comprises a nucleotide sequence which confers antibiotic resistance in a host cell population. Such markers include amikacin, augmentin (amoxicillin plus clavulonic acid), ampicillin, cefazolin, cefoxitin, ceftazidime, ceftiofur, cephalothin, enrofloxacin, florfenicol, gentamicin, imipenem, kanamycin, penicillin, sarafloxicin, spectinomycin, streptomycin, tetracycline, ticarcillin, tilmicosin, or chloramphenicol resistance genes. Other suitable markers will be readily apparent to the skilled artisan.

The invention also provides for a method of producing a host cell where the expression vectors of the current invention have been introduced into the host cell. Methods of introducing genetic material into host cells, such as those described in typical molecular biology laboratory manuals, for example J. Sambrook, E.F. Fritsch and T. Maniatis, *Molecular Cloning: A Laboratory Manual*, 2d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1989), are well known to the skilled artisan. These methods include, but are not limited to, calcium phosphate transfection, DEAE-dextran mediated transfection, microinjection, lipid-mediated transfection, electroporation or infection. Accordingly, a preferred embodiment of the present invention provides a host cell comprising the vector of the present invention.

eukaryotic cell where the desired nucleic acid sequence has been introduced into the cell. There are a variety of suitable host cells, including but not limited to bacterial, fungal, insect, mammalian and plant cells, that can be utilized in the present invention. Representative bacterial host cells include, but are not limited to, *Streptococci, Staphylococci, E. coli, Streptomyces, Bacillus* and *Corynebacterium*. Representative fungal cells include but are not limited to, yeast cells and *Aspergillus*. Insect cells include, but are not limited to, *Drosophila* S2 and *Spodoptera* Sf9 cells. Examples of mammalian cells include, but are not limited to, Drosophila S2 and Spodoptera Sf9 cells. Examples of mammalian cells include, but are not limited to, CHO, COS and Hela cells.

The present invention provides methods for utilizing the nucleic acid of [0035] SEQ ID NO:1 or SEQ ID NO:3, which encodes the amino acid sequence of a mutant pyruvate carboxylase. Such methods include the replacement of the wildtype pyruvate carboxylase with the feedback-resistant pyruvate carboxylase, and the production of amino acids. The method for replacement of a wild-type pyruvate carboxylase gene, with a feedback resistant pyruvate carboxylase gene, in a Corynebacterium glutamicum host cell comprises the steps of: (a) replacing a genomic copy of the wild-type pyruvate carboxylase gene with a selectable marker gene through homologous recombination to form a first recombinant strain; and (b) replacing the selectable marker gene of step (a) in the first recombinant strain, with the feedback resistant pyruvate carboxylase gene through homologous recombination to form a second recombinant strain. The homologous recombination in steps (a) and (b) would occur between the genetic material of the host cell and any of the vectors of the present invention.

[0036] Homologous recombination is a technique that is used to disrupt endogenous nucleotide sequences in a host cell. Normally, when an exogenous nucleotide sequence is inserted into a host cell, this polynucleotide may randomly insert into any area of the host cell's genome, including endogenous plasmids. However, with homologous recombination, the exogenous nucleotide

sequence contains sequences that are homologous to an endogenous nucleotide sequence within the host cell. Once introduced into the cell, for example by electroporation, the exogenous nucleotide sequence will preferentially recombine with and replace the endogenous nucleotide sequence with which it is homologous.

Sequence which is not found in the host cell. Thus, the term exogenous nucleotide sequence is meant to encompass a nucleotide sequence that is foreign to the host cell, as well as a nucleotide sequence endogenous, or native, to the host cell that has been modified. Modification of the endogenous nucleotide sequence may include, for instance, mutation of the native nucleotide sequence or any of its regulatory elements. As used herein, mutation is defined as any change in the wild-type sequence of the host's genetic material, including plasmid DNA. An additional form of modification may also include fusion of the endogenous nucleotide sequence to a nucleotide sequence that is normally not present, in relation to the endogenous nucleotide sequence.

[0038] Host cells that have undergone homologous recombination are selected on the basis of antibiotic resistance through the use of, for example, the selectable markers mentioned above. The process of selecting cells that have undergone homologous recombination will be readily apparent to one skilled in the art.

Another aspect of the current invention is a method for producing amino acids. In the current context, production of amino acids is accomplished by culturing host cells where a vector of the present invention has been introduced into the host cell, or culturing host cells where homologous recombination, involving a vector of the present invention, has taken place. Culturing of the host cells is performed in the appropriate culture media. Subsequent to culturing the host cells in culture media, the desired amino acids are separated from the culture media. Preferably, the amino acids produced by the methods described herein include L-lysine, L-threonine, L-methionine, L-isoleucine, L-glutamate, L-